

SAMPLE PREPARATION

Make sure Strips, Buffer and water are at room temperature before testing

Make sure you have scanned the Multi-Matrix Barcode Card (MMBC) that matches your kit lot

Set Incubator at 22°C

You will need a 100 µL pipette



1. Collect a composite DDGS sample according to your own sampling plan or USDA/GIPSA guidelines



2. Grind samples to pass through a 20 mesh sieve. Mix ground material thoroughly before sub-sampling. Carefully weigh out 20-50g subsample.



3. Add 2.5X of 50% ethanol (20g sample requires 50 mL; 50g sample requires 125 mL)



4. Cover and shake on a mechanical shaker at highest speed for 1 minute (or vigorously by hand, 2 min.)



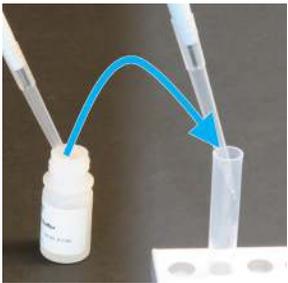
5. Take a portion of extract and centrifuge for 30 seconds at 2000 x g (not RPM) - follow manufacturer's instructions

TEST PROCEDURE

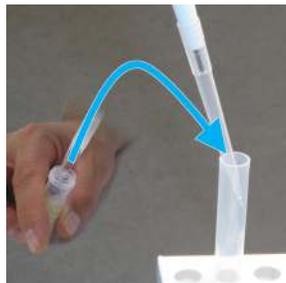
(more detailed instructions in the Product Insert)

Ensure Incubator has reached 22°C

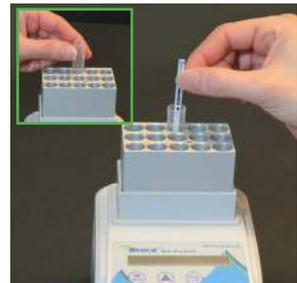
*Note: acclimation is only required when the ambient temperature is unknown or outside of 20-24°C (68-75°F)



6. Add 100 µL Buffer to a reaction tube



7. Transfer 100 µL clarified extract to the tube; mix well with pipette tip (stir or pipette up and down)



8. Place tube in Incubator and acclimate for 2 min.*, then add QuickTox Strip; wait 5 minutes for results

QuickScan TEST RESULTS

(more detailed instructions in the QuickScan User Manual)

9. Remove strip from vial immediately after the 5 minute test time. Cut off and discard bottom pad with arrow tape. (No drying step!)

10. Place in the QuickScan carrier and slide carrier in. Click "Read Test" on Main Menu. Results are reported between 100 and 1000 ppb.

11. Results Screen will appear when scanning is complete. Enter sample identification data and use buttons to save or print report.